

Preliminary communication

Conformation of the pentasaccharide corresponding to the binding site of heparin to Antithrombin-III

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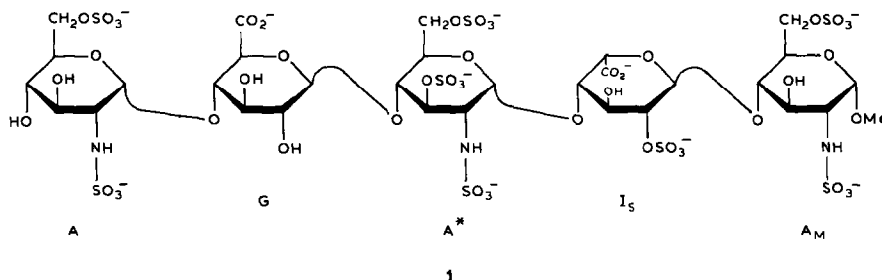
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The minimal binding site of heparin to Antithrombin-III (AT-III) is represented by a pentasaccharide¹⁻² which was recently synthesised³. We now propose a model for the synthetic pentasaccharide⁴ **1** based on force-field computations and n.m.r. (essentially n.O.e.) measurements.



A major problem is the determination of the conformation of the iduronate residue I_s which has been a matter of controversy^{5c,6}. Force-field computations indicated that, in addition to the 1C_4 and 4C_1 conformations, a third almost equi-energetic conformation, the skew boat 2S_0 , must be considered for the iduronate ring^{5a}.

The ${}^3J_{H,H}$ values for the iduronate residue in various mono- and oligo-saccharides related to the heparin binding site for AT-III have been interpreted in

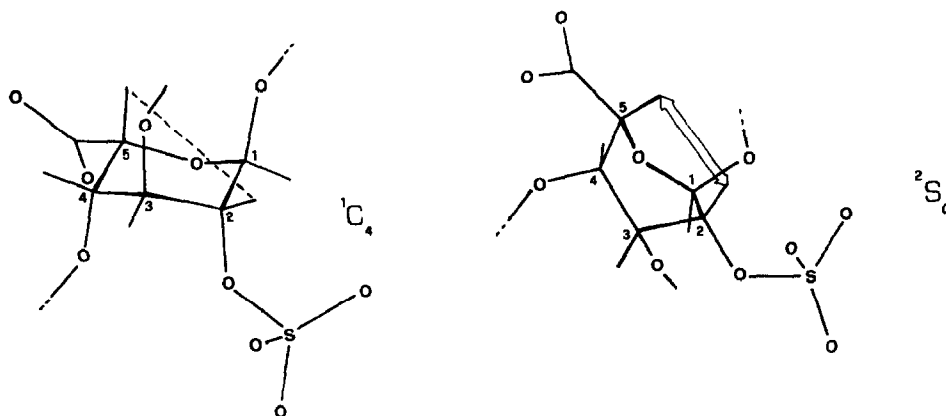


Fig. 1. The two conformations of I_5 observed in heparin and inside oligosaccharide sequences. The short H-2/H-5 distance in the 2S_0 form is indicated.

terms of conformer populations^{5d}, the relative abundance of which is governed both by the sequence of monosaccharide units and by the pattern of substitution^{5b,5d}. When present as an internal unit in an oligosaccharide, the iduronate residue is in equilibrium between the 1C_4 and 2S_0 conformations (Fig. 1).

Force-field computations carried out on **1** provided several models, and the procedure outlined below led to the selection of the most probable.

The φ, ψ energy maps were inspected first for the four pairs of interglycosidic torsional angles. Taking into account the three possible conformations of the iduronate ring, but neglecting differences in the side-chain conformations, sixteen conformers of the pentasaccharide were found with comparable energy (within 3.5 kcal.mol⁻¹).

Analysis of the $J_{H,H}$ values for **1** indicates no appreciable amount of the 4C_1 conformation of the iduronate ring^{5d}; hence, the corresponding conformers were discarded. Furthermore, by restricting the energy range to 2 kcal.mol⁻¹, a series of conformers, characterised by the values -7° and 39° for φ and ψ in the A-G interglycosidic linkage, can also be discarded.

Only the four remaining conformers were then considered; these corresponded to the 1C_4 and 2S_0 conformations for I_5 , combined with the two possible φ, ψ angles for the I_5-A_M interglycosidic linkage. The values of φ and ψ for the four conformers are given in Table I, and the two most stable are shown in Fig. 2.

Using 500-MHz n.m.r. spectroscopy, both intra- and inter-residue negative n.O.e. effects were observed at 10° on a phase-sensitive NOESY map⁷ (Fig. 3) and quantified by mono-dimensional n.O.e. difference spectroscopy. The n.O.e. observed between H-1 and H-2 of the three 2-amino-2-deoxyglucose residues was essentially the same (-16 to -17%), indicating isotropic tumbling. Particularly noticeable is the strong n.O.e. effect [-13% , *i.e.*, only slightly smaller than those (17 – 20%) observed for vicinal diequatorial protons] between H-5 and H-2 of residue I_5 , as predicted for a prevalent 2S_0 conformation by the above calculations.

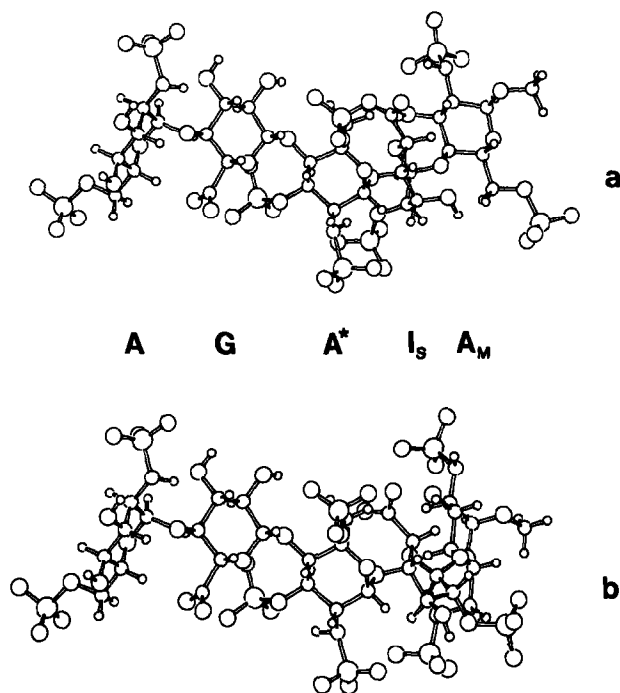


Fig. 2. The two most stable conformers computed for **1**: (a) minimum c I (according to Table I), corresponding to the 1C_4 conformation of I_s; (b) minimum s II (conformation 2S_0).

TABLE I

INTERGLYCOSIDIC DIHEDRAL ANGLES $\varphi(H-1-C-1)O-1-C-4$ AND $\psi(C-1-O-1)C-4-H-4$ IN THE FOUR MORE STABLE CONFORMERS OF **1**, AND THEIR RELATIVE ENERGIES COMPUTED ACCORDING TO REFS. 5a AND 5d

Minimum ^a	$(\varphi_1; \psi_1)$	$(\varphi_2; \psi_2)$	$(\varphi_3; \psi_3)$	$(\varphi_4; \psi_4)$	$E (kcal.mol^{-1})$
c I	-36; -26	49; 4	-37; -33	24; -49	0.31
c II	-36; -26	49; 4	-35; -31	31; 19	1.34
s I	-36; -26	50; 4	-35; -41	34; -43	1.27
s II	-36; -27	48; 5	-38; -44	44; 9	0

^aMinima c correspond to the 1C_4 conformation of I_s, whereas s refers to 2S_0 .

Indeed, in the skew-boat form, the H-2 to H-5 distance is the shortest interproton distance (2.3 Å, as compared with 2.8 Å for H-1 to H-3), whereas in the chair form it is 4.0 Å (see Fig. 1). This provides support for the occurrence of this conformer.

The present picture of the average conformation has the following features. (a) The A-G-A* sequence is quite rigid, owing to a single conformation of the glycosidic linkage. The present calculations indicate that the rigidity of the G-A* linkage arises from the β -D-glucuronate residue and from the interactions of the peculiar 3-O-sulfo group with O-5 and COO⁻ of G. (b) In the A*-I-A_M sequence

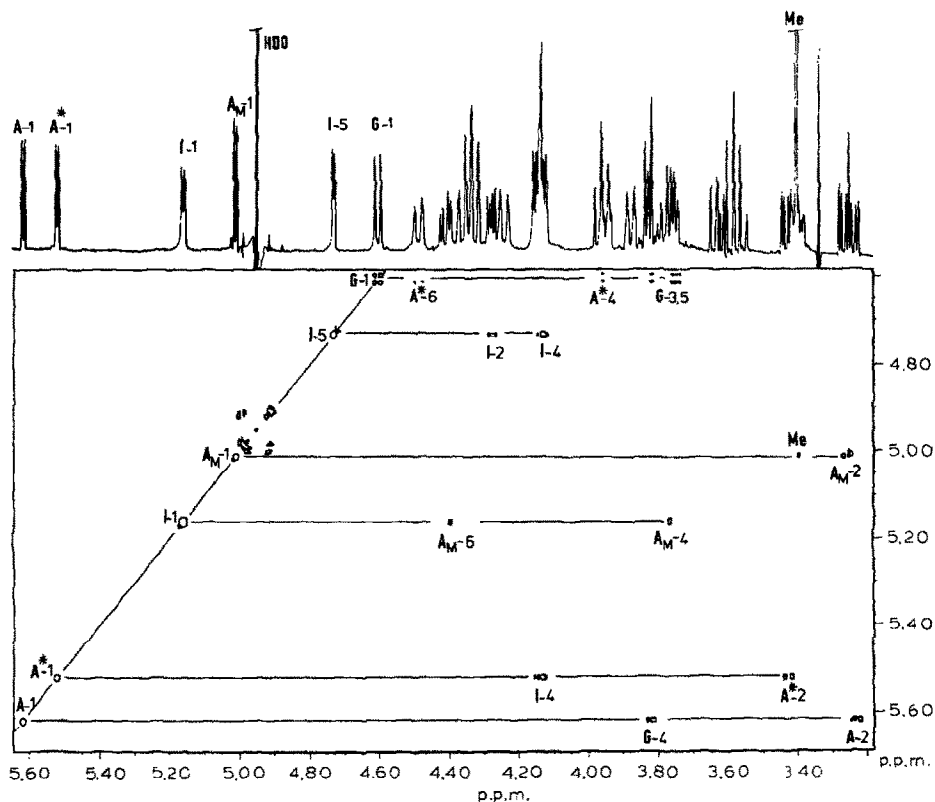


Fig. 3. 500-MHz phase-sensitive NOESY map (partial) of AGA*IA_M in ²H₂O at 20°. Only negative contours have been plotted.

(in the four conformers), the iduronate ring is in equilibrium between two conformations, the relative populations computed from the $^3J_{\text{HH}}$ values being $^2S_0 \sim 60\%$ and $^1C_4 \sim 40\%$; the presence of the 2S_0 form is confirmed by the marked n.O.e. effect.

Thus, four main conformers are proposed as possible candidates for the pentasaccharide **1** in solution corresponding to the binding site to antithrombin. A more refined force-field and n.O.e. study is in progress to improve this model.

REFERENCES

- 1 J. CHOAY, J.-C. LORMEAU, M. PETITOU, P. SINAY, AND J. FAREED, *Ann. N.Y. Acad. Sci.*, 370 (1981) 644-649.
- 2 L. THUNBERG, G. BÄCKSTRÖM, AND U. LINDAHL, *Carbohydr. Res.*, 100 (1982) 393-410.
- 3 J. CHOAY, M. PETITOU, J.-C. LORMEAU, P. SINAY, B. CASU, AND G. GATTI, *Biochem. Biophys. Res. Commun.*, 116 (1983) 492-499; P. SINAY, J.-C. JACQUINET, M. PETITOU, P. DUCHAUSSOY, I. LEDERMAN, J. CHOAY, AND G. TORRI, *Carbohydr. Res.*, 132 (1984) c5-c9; C. A. A. VAN BOECKEL, T. BEETZ, J. N. VOS, A. H. M. DE JONG, S. F. VAN AELST, R. H. VAN DEN BOSCH, J. M. R. MERTEENS.

- AND F. A. VAN DER VLUGT, *J. Carbohydr. Chem.*, 4 (1985) 293–321; Y. ICHIKAWA, R. MONDEN, AND H. KUZUHARA, *Tetrahedron Lett.*, (1986) 611–614; M. PETITOU, P. DUCHAUSSOY, I. LEDERMAN, J. CHOAY, P. SINAÏ, J.-C. JACQUINET, AND G. TORRI, *Carbohydr. Res.*, 147 (1986) 221–236.
- 4 M. PETITOU, P. DUCHAUSSOY, I. LEDERMAN, J. CHOAY, J.-C. JACQUINET, P. SINAÏ, AND G. TORRI, *Carbohydr. Res.*, in press.
- 5 (a) M. RAGAZZI, D. R. FERRO, AND A. PROVASOLI, *J. Comput. Chem.*, 7 (1986) 105–112; (b) G. TORRI, B. CASU, G. GATTI, M. PETITOU, J. CHOAY, J.-C. JACQUINET, AND P. SINAÏ, *Biochem. Biophys. Res. Commun.*, 128 (1985) 134–140; (c) B. CASU, J. CHOAY, D. R. FERRO, G. GATTI, J.-C. JACQUINET, M. PETITOU, A. PROVASOLI, M. RAGAZZI, P. SINAÏ, AND G. TORRI, *Nature (London)*, 322 (1986) 215–216; (d) D. R. FERRO, A. PROVASOLI, M. RAGAZZI, G. TORRI, B. CASU, G. GATTI, J.-C. JACQUINET, P. SINAÏ, M. PETITOU, AND J. CHOAY, *J. Am. Chem. Soc.*, 108 (1986) 6773–6778.
- 6 D. A. REES, E. R. MORRIS, J. F. STODDART, AND E. S. STEVENS, *Nature (London)*, 317 (1985) 480.
- 7 G. BODENHAUSEN, H. KOGLER, AND R. R. ERNST, *J. Magn. Reson.*, 58 (1984) 370–388.